

Variable virulence in Atlantic salmon of an ISAV isolate in repeated experimental challenges and Lack of protective effects in an mRNA vaccine against ISA

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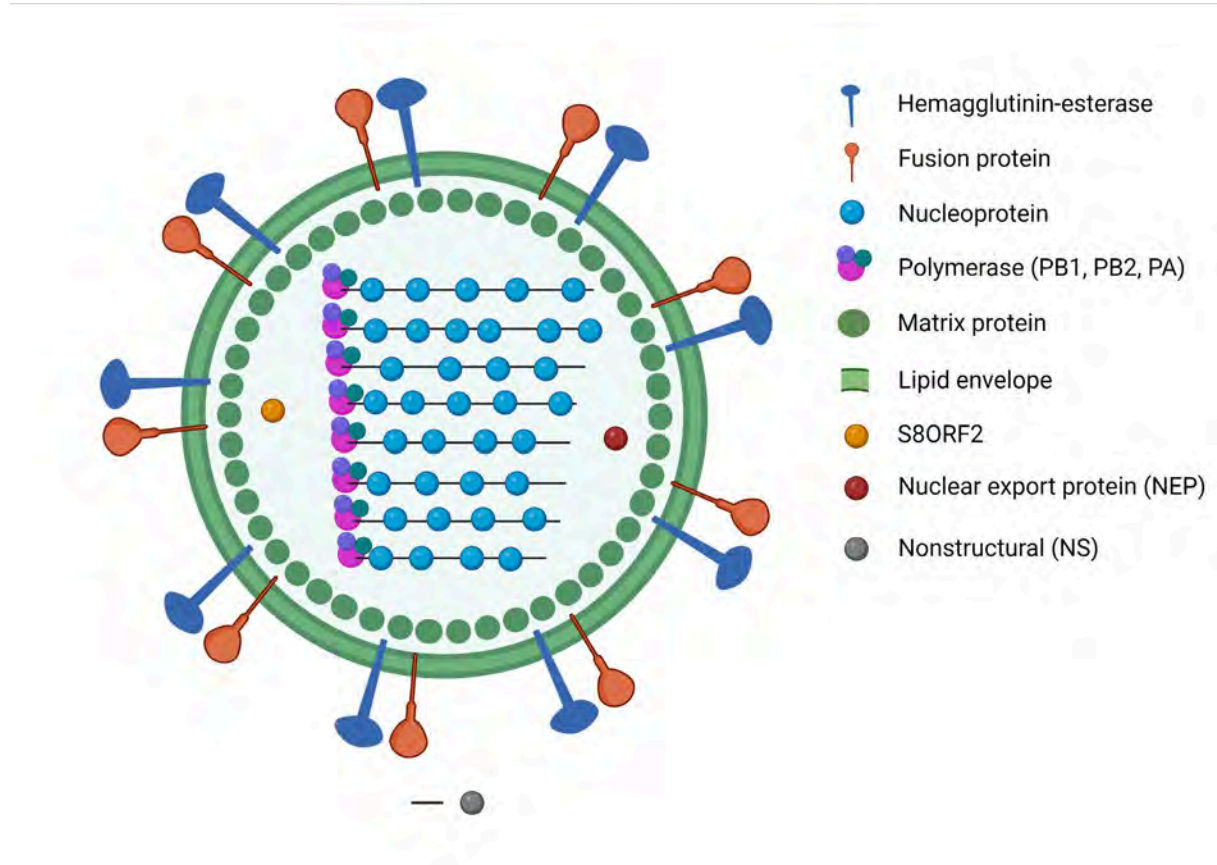
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Introduction

Infectious salmon anemia virus (ISAV)

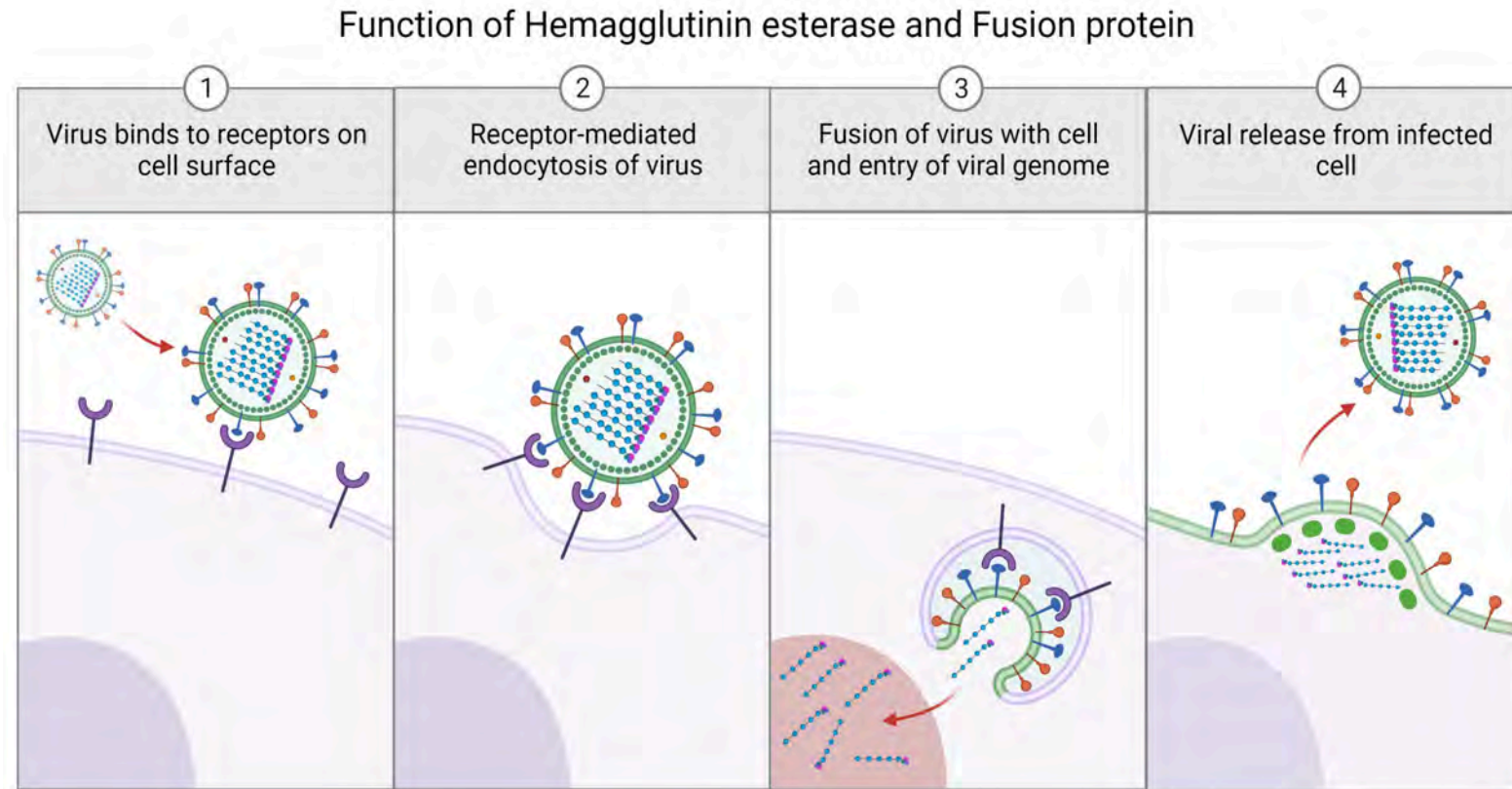
- Segmented RNA genome
- 8 segments encoding 10 proteins
- Current ISAV vaccines may be assessed as not optimal

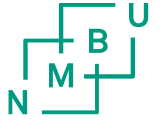


Introduction

Hemagglutinin esterase and Fusion protein

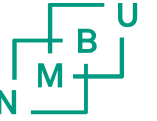
- 2 surface glycoproteins:
 - Hemagglutinin esterase (HE)
 - Receptor binding and destroying activity
 - Fusion (F) protein
 - Viral fusion
- Vaccines should target these surface proteins





mRNA vaccination method

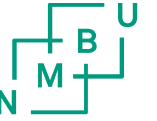
- Generate mRNA and encapsulate them in lipid nanoparticles (LNP)
- Vaccination by intramuscular injection of Atlantic salmon
- ISAV challenge
- Mortality monitoring
- Classical ISAV challenges usually uses a well characterized high virulent virus isolate
- In this experiment we wanted to use a recent isolate – assumed to be less virulent than the classical ones



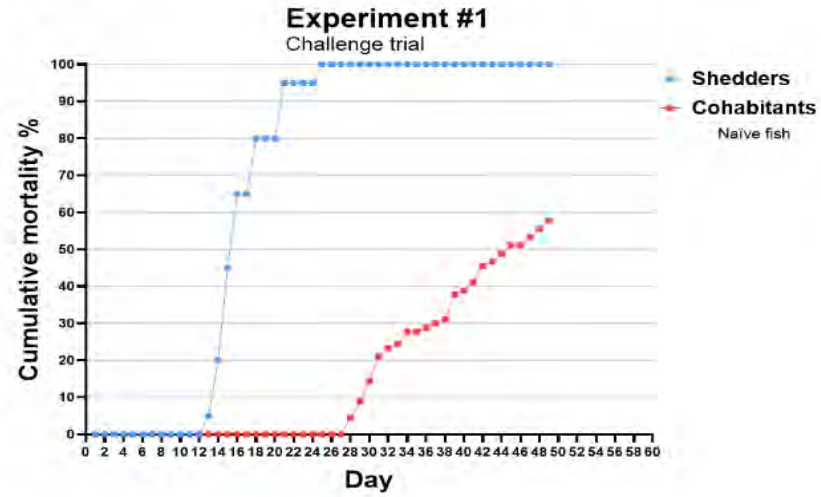
Observed variable virulence of same ISAV isolate in repeated experiments

- Not all experiments go as planned...
- Originally, the experiments were conducted to assess vaccine efficacy of an mRNA vaccine against ISA using a recent virus isolate from 2020
- Four separate viral challenge trials were performed using the same virus isolate
- However, vaccine efficacy was not concludable due to lack of viral infection in the vaccination challenges
- Therefore, we sought to investigate plausible reasons for the lack of infection between challenge trials

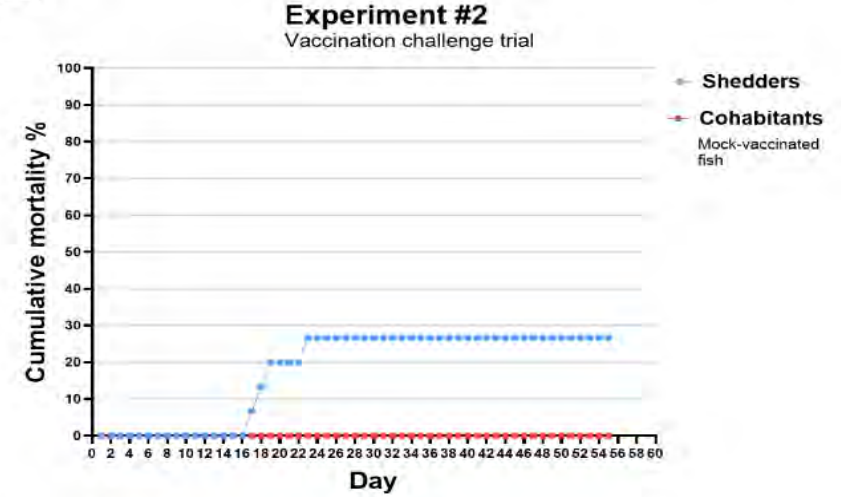
Mortality data



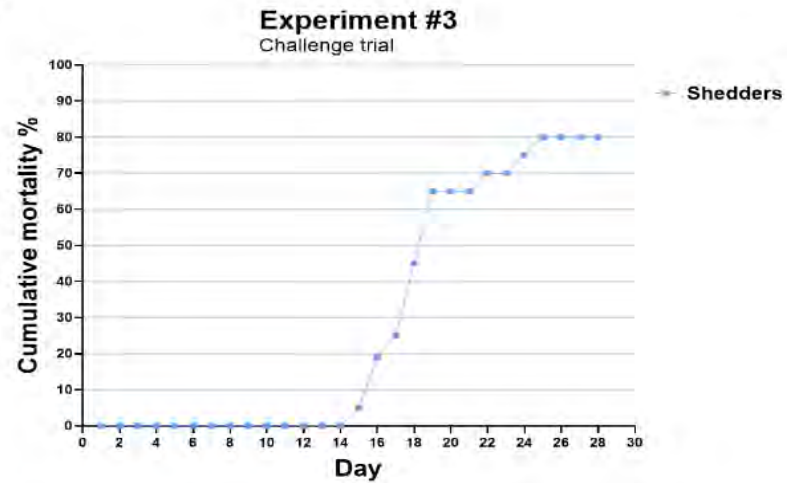
A



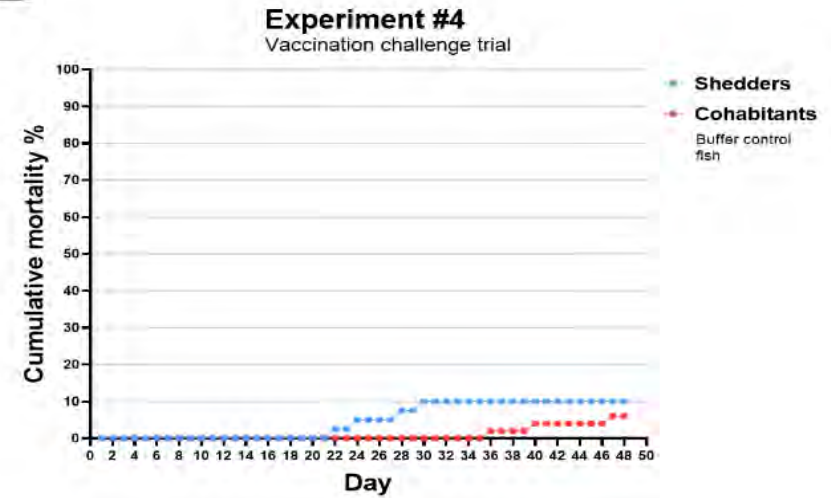
B



C



D

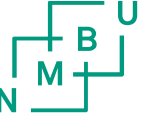


Overview of parameters in all challenge experiments



- Experiment 1 and 2 were challenged with same ISAV batch
- Experiment 3 and 4 were challenged with another ISAV batch
- Viral titer was unaffected after months of storage
- Factors that may have affected shedders:
 - Fish density
 - Tank size

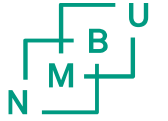
Conditions	Experiment #1 Challenge trial	Experiment #2 Vaccination challenge trial	Experiment #3 Challenge trial	Experiment #4 Vaccination challenge trial
Species	Atlantic salmon (<i>Salmo salar</i>)			
Strain	Stofnfiskur			
Origin	Veso Aqualab Hatchery			
Average weight	25-40 grams			
Physiological status	Parr			
Inoculum	Virus isolate: 70-72			
	Amount of infectious virus per fish: 10 ⁴ TCID ₅₀			
Number of shedder fish	20	15	20	40
Number of cohabitant fish*	90	50	-	50
Total number of fish	110	165*	20	240**
Cohabitant status	Naïve	Mock-vaccinated	-	Tris-HCl buffer injected
Tank size	130L	450L	130L	450L
Fish density***	27.5 g/L	11.9 g/L	5 g/L	17.3 g/L
Shedder density per volume	0.15 fish/L	0.03 fish/L	0.15 fish/L	0.09 fish/L
Salinity	Fresh water			
Stocking density	Max 50 kg/m ³			
Temperature	12°C ± 1°C			
Flow	0.8 l/kg fish per minute			
Water discharge	Tube overflow system			
Cleaning	Once a day			
Photoperiod regime	L:D = 24:0			
Feeding	Automatic feeders, 1% biomass per day			



Conclusion?

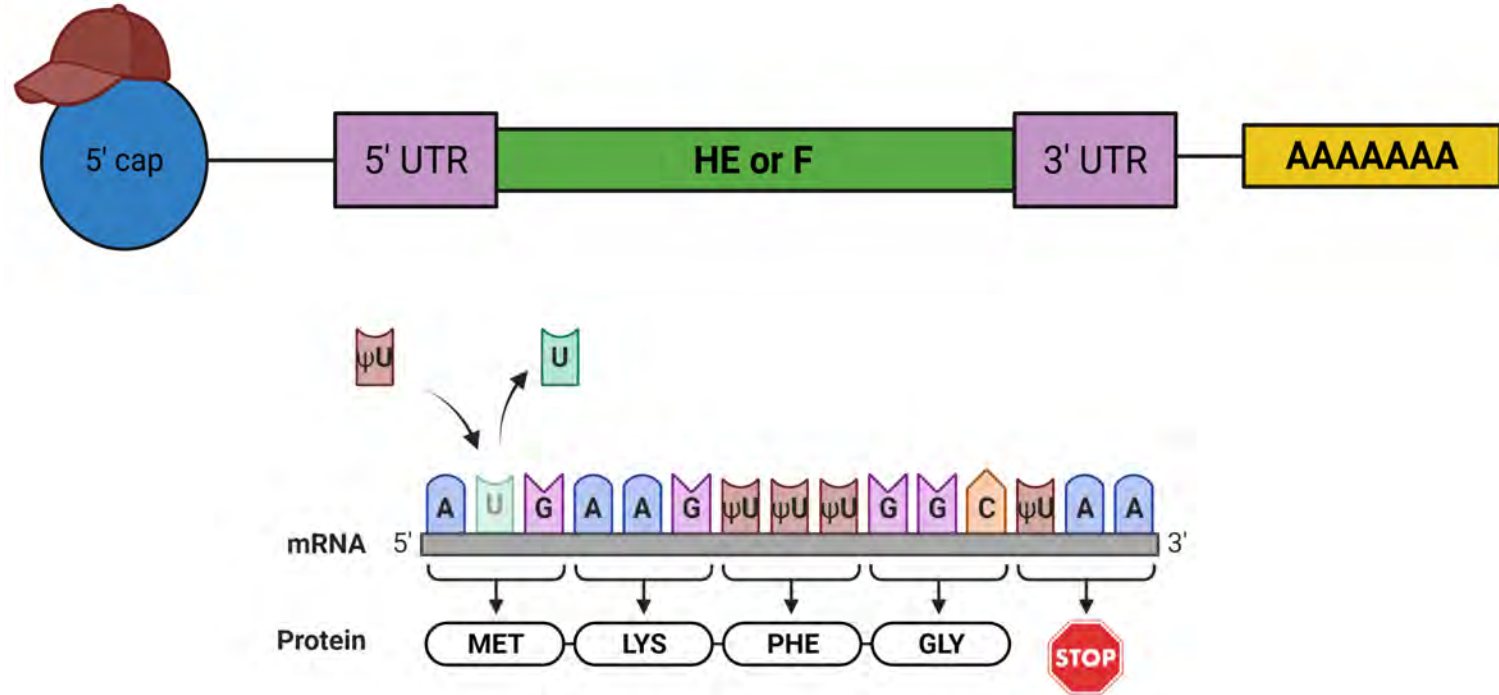
- Predicting virulence of uncharacterized ISAV isolates proves challenging
- Slight changes in design of challenge experiments might have considerably impact on observed virulence of assumed "low" virulent isolates
- Research of zebrafish observed elevated stress levels in small tanks compared to larger tanks (unaffected by fish densities within reasonable limits)

Introduction mRNA vaccine technology



• Construction of mRNA:

- Enzymatical addition of 5' Cap
- Poly-A tail
- Untranslated regions (UTRs)
- Substitution of N1-methyl pseudouridine (COVID)
 - CureVac → 40%
 - Moderna/Pfizer-BioNTech → 95%



Introduction mRNA vaccine technology

- **Lipid nanoparticles (LNPs) for mRNA delivery**

- Consists of 4 different lipids

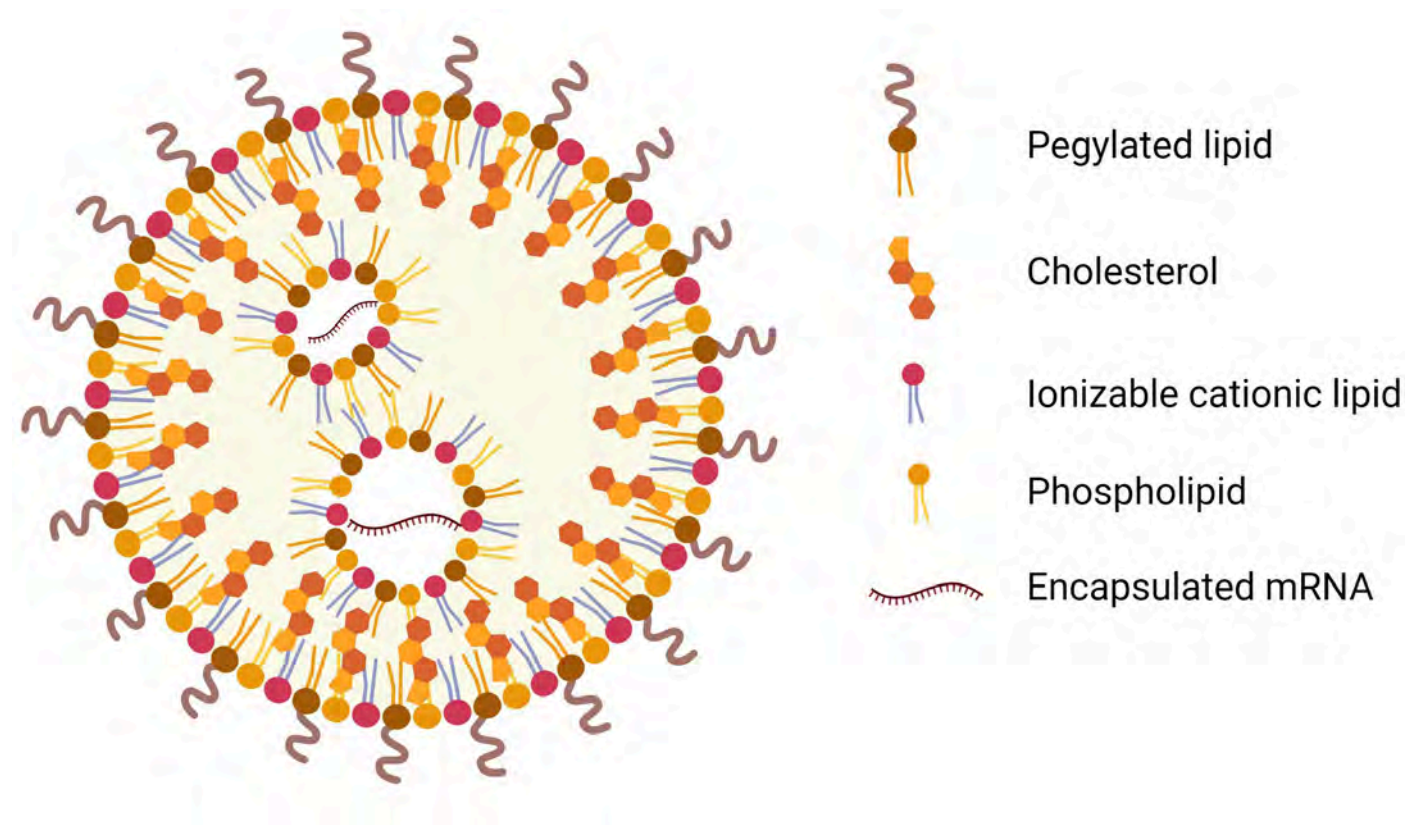
- Ionizable lipids

- "Helper lipids"

- Cholesterol

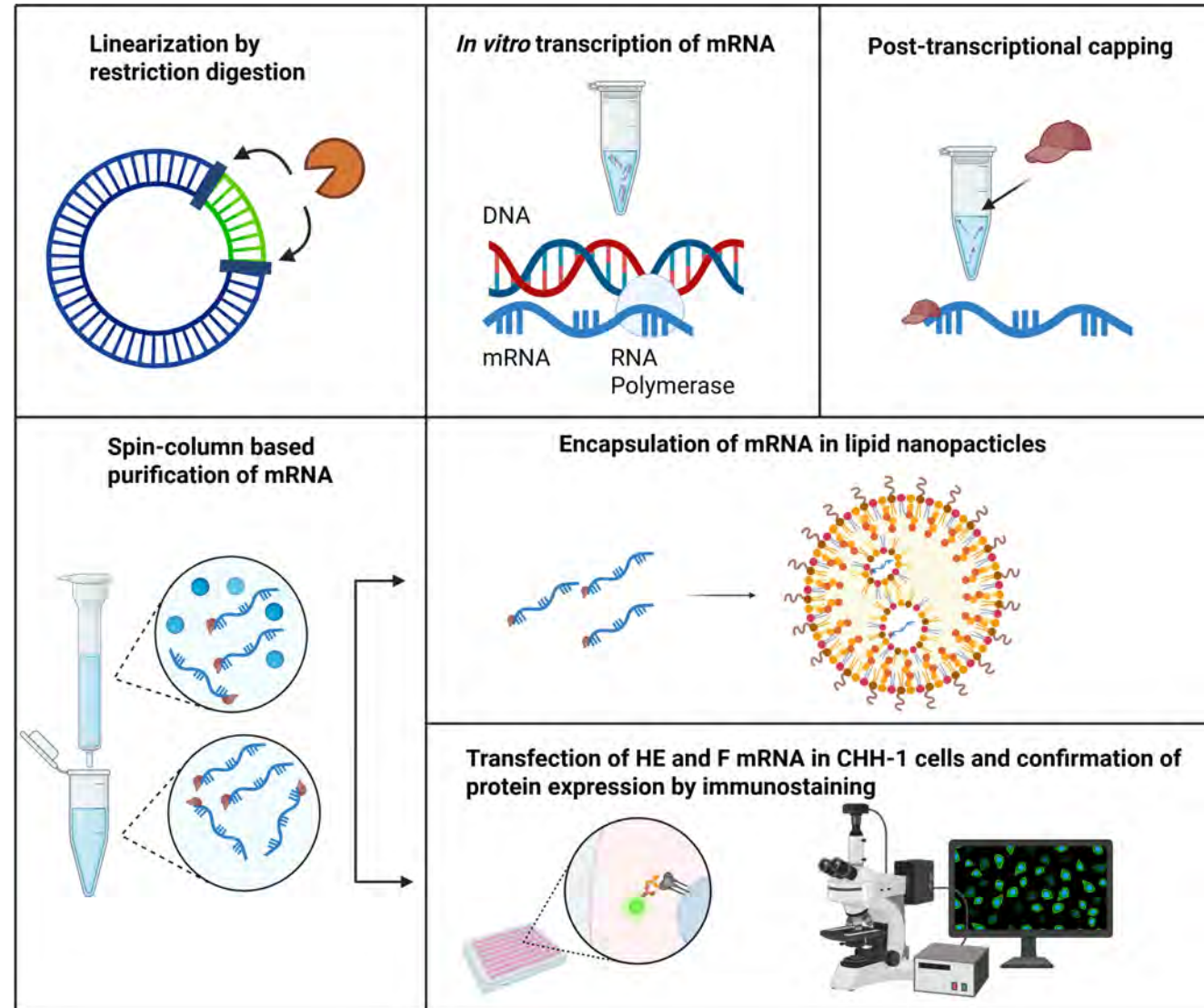
- Phospholipids

- PEGylated lipids



Generation of mRNA-LNPs (vaccine)

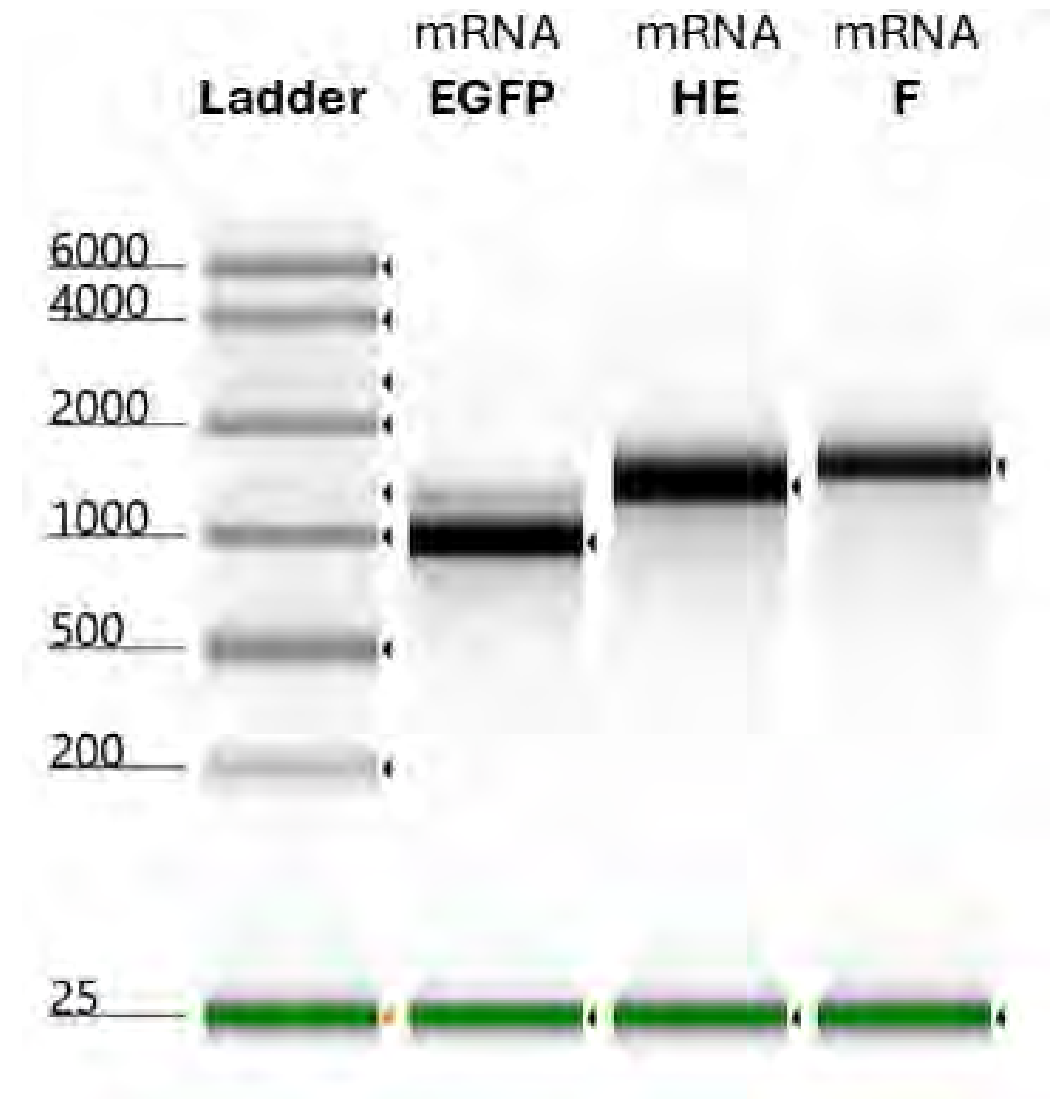
- Design of DNA plasmids encoding HE or F containing a 120 Poly(A) tail (also included EGFP mRNA as control)
- *In vitro* transcription of mRNAs with N1-methyl-pseudouridine
- Post-transcriptional capping, including Cap0 (guanosine-7-methyl) and Cap1 (methylation of the 2'-O position of the first transcribed nucleotide)
- Immunostaining of mRNA-transfected CHH-1 cells to confirm HE and F protein expression
- LNP encapsulation of mRNA by SINTEF (similar composition as Moderna's LNPs)





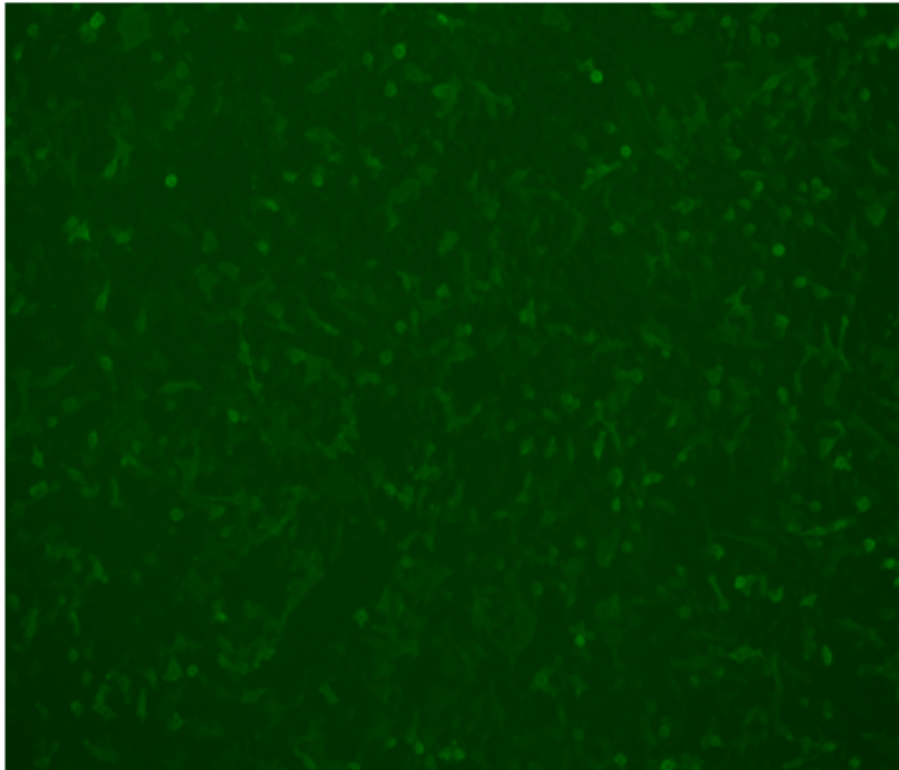
mRNA construction

- Confirmation of mRNA sizes by capillary gel electrophoresis
- Theoretical size EGFP: 988 nt
- Theoretical size HE: 1444 nt
- Theoretical size F: 1609 nt

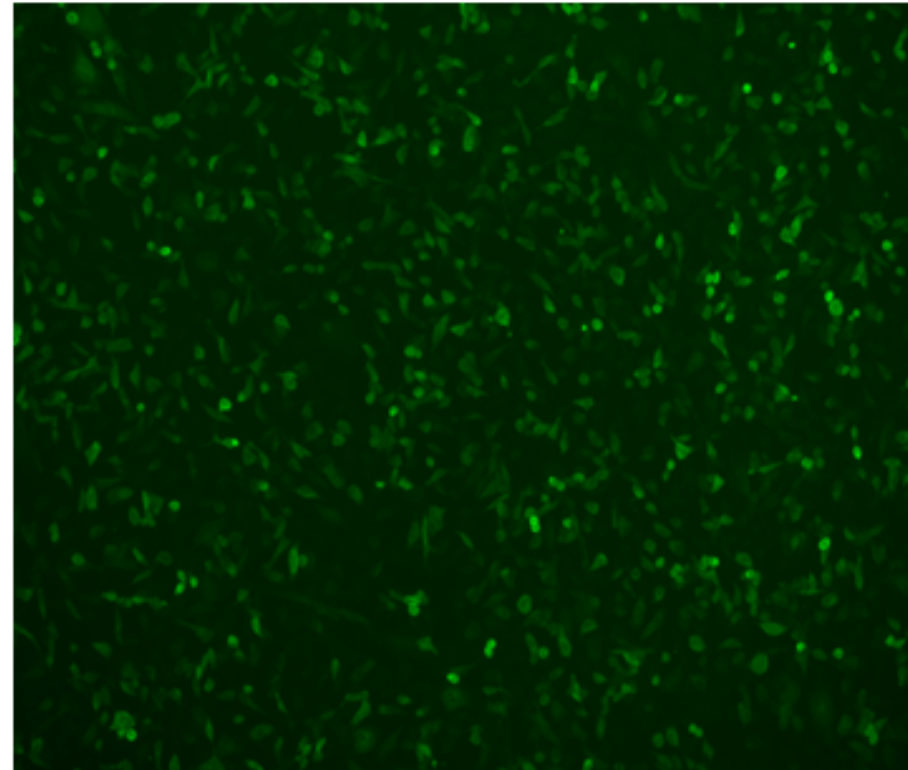


Immunostaining of CHH-1 cells transfected with HE or F mRNA (3 days post transfection)

HE mRNA



F mRNA



Study design



70 fish per group per tank

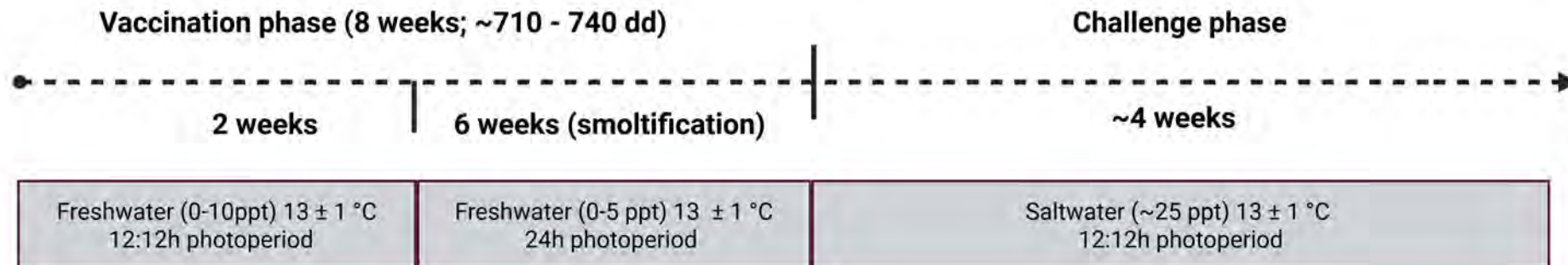
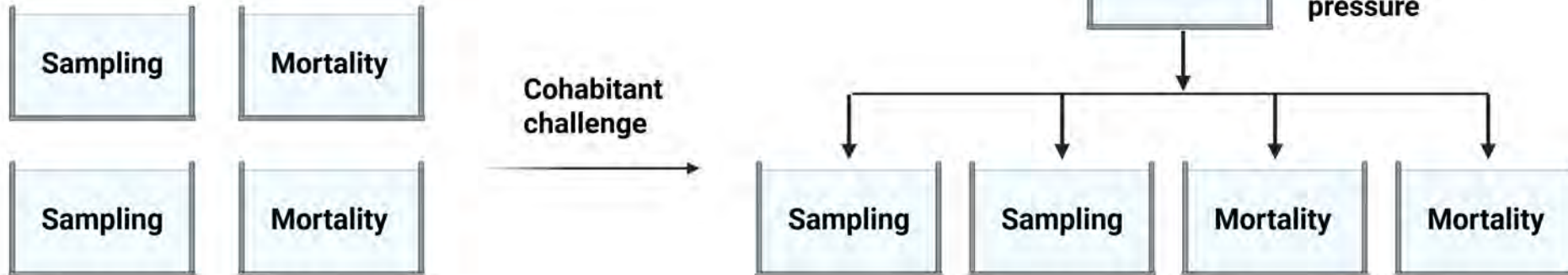
Group 1: 10µg HE

Group 2: 10 µg F

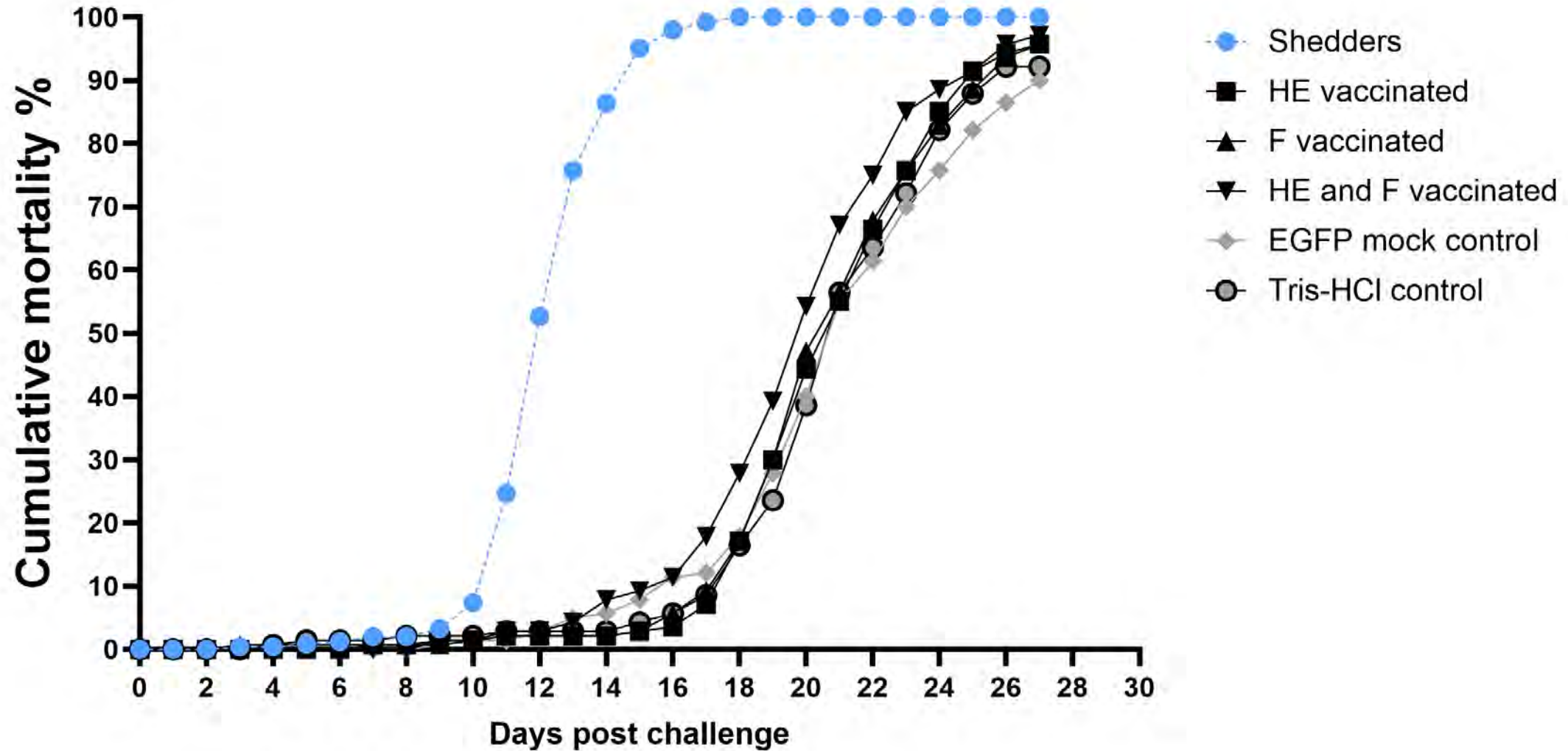
Group 3: 10 µg HE and 10 µg F

Group 4: 10 µg EGFP mock control

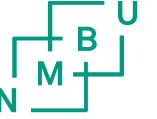
Group 5: Tris-HCl control



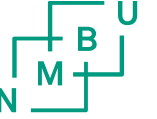
Mortality data from vaccine experiment



Notes



- Difference between ISAV challenge strain and mRNA sequence (NA-HPR4 and Glesvær)
- While protein expression was confirmed after IVT of mRNA, this was not checked after LNP formulation or after arrival at the fish research facility
 - LNP-encapsulated EGFP demonstrated protein expression both *in vitro* and *in vivo*
 - Vaccine shipment was still in a frozen condition at receipt



Conclusion

- Our mRNA–LNP vaccines did not appear to prevent infection of ISAV
- Studies in rainbow trout have shown protection against rhabdovirus infections when administering mRNA vaccines
- Future studies should aim to optimize this vaccine technology for Atlantic salmon, including LNP formulation, mRNA construct optimization, administration route and dosage, self-amplifying mRNA(?)



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